

The role of VE-cadherin in vascular permeability

Commentary on Elisabetta Dejana's paper published on Nature Communications

by Michael Gimbrone

The vascular endothelium, the single-cell-thick lining of the human cardiovascular system, is a living membrane that comprises a container for blood. The integrity of this container is essential for the health of the various tissues and organs of the body, and changes in its permeability are a hallmark of acute and chronic inflammatory disease processes.

The building blocks of this dynamic barrier are individual endothelial cells, which are linked together by a complex of intercellular molecular bridges that serve as the "gate-keepers" for the trans-endothelial movement of proteins, fluid, and cells.

For many years it has been appreciated that this endothelial barrier function varies in the different types of vessels-arteries, veins, capillariesthat comprise the branching vascular tree. Interestingly, inflammatory mediators, such as histamine and bradykinin, show a selective effect on the venous side of the microcirculation. The IFOM research group led by Professor Elisabetta Dejana has been at the forefront of dissecting the molecular components of these endothelial junctional complexes, and the mechanisms of their regulation in health and disease. In this report, they provide an in-depth analysis of the biochemical processes controlling the cell-cell "zipper" function of VE-cadherin, a key determinant of endothelial permeability, in the context of the inflammatory response.

Specifically, they establish that phosphorylation of certain amino acids within the structure of VEcadherin occurs constitutively within veins, but not arteries, in vivo, and that inflammatory mediators such as bradykinin, preferentially act to enhance the internalization/degradation of these phosphorylated junctional proteins, resulting in a localized, rapid but transient, increase in vascular permeability. In effect, this provides a mechanistic explanation for the topography of increased permeability in inflammation. Further, they establish that differences in the magnitude of the fluid mechanical forces imparted by blood flow (lower in veins than in arteries) may be responsible for the observed spatial pattern of VE-cadherin phosphorylation that sensitizes vessels to the action of inflammatory mediators.

A major strength of this work is the skillful use of both in vitro (cultured cell models) and in vivo (intravital molecular microscopy) strategies to probe the cellular and molecular mechanisms of these fundamental vascular biological processes. Their novel results point the way to potential new pharmacologic interventions in inflammatory diseases. Phosphorylation of VE-cadherin is modulated by haemodynamic forces and contributes to the regulation of vascular permeability in vivo.

Endothelial adherens junctions maintain vascular integrity. Arteries and veins differ in their permeability but whether organization and strength of their adherens junctions vary has not been demonstrated in vivo. Here we report that vascular endothelial cadherin, an endothelial specific adhesion protein located at adherens junctions, is phosphorylated in Y658 and Y685 in vivo in veins but not in arteries under resting conditions. This difference is due to shear stress-induced junctional Src activation in veins. Phosphorylated vascular endothelial-cadherin is internalized and ubiquitinated in response to permeabilityincreasing agents such as bradykinin and histamine. Inhibition of Src blocks vascular endothelial cadherin phosphorylation and bradykinin-induced permeability. Point mutation of Y658F and Y685F prevents vascular endothelial cadherin internalization, ubiquitination and an increase in permeability by bradykinin in vitro. Thus, phosphorylation of vascular endothelial cadherin contributes to a dynamic state of adherens junctions, but is not sufficient to increase vascular permeability in the absence of inflammatory agents. [PMID: 23169049]









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